



The sticky–Slide family allows you to perform cell culture experiments with custom–specific bottom materials like plastic sheets, glass slides, spotted coverslips, printed circuit boards, etc. The self adhesive ("sticky") underside of the bottomless blank slide is easily adapted to your specific substrate by pressing on by hand.

The sticky–Slide 8 Well provides a common open well format which is best suited for maximum sample access in a wide variety of experimental applications.

Material

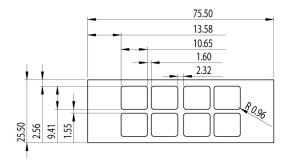
The slide material of sticky–Slides is identical to common μ –Slides (uncoated). The Slides are not autoclavable since they are temperature stable up to $60^{\circ}\text{C}/140^{\circ}\text{F}$ only. All sticky–Slides are delivered sterile and single packed. Please keep in mind that sterility is lost when non–sterile substrates are used.

The sticky bottom itself is a 50 µm biocompatible double–faced adhesive tape. The tape is covered by a protection film which has to be removed before usage.

Geometry

All technical details beside bottom material are identical to μ -Slide 8 Well. The Slides provide standard slide format according to ISO 8037/1.

| Geometry of the sticky-Slide 8 Well | | |
|--|--------------------------|--|
| Number of wells | 8 | |
| Dimension of wells $(w \times l \times h)$ in mm | $9.4\times10.7\times6.8$ | |
| Recommended volume per well | 300 µl | |
| Total height with lid | 8 mm | |
| Growth area per well | 1.0 cm^2 | |
| Bottom | none | |



Shipping and Storage

The μ –Slides, μ –Dishes and μ –Plates are sterilized and welded in a gas-permeable packaging. The shelf life under

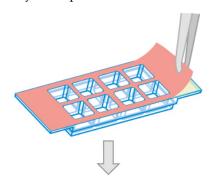
proper storage conditions (in a dry place, no direct sunlight) is listed in the following table.

| Conditions | | |
|----------------------------------|--------------|--|
| Shipping conditions | Ambient | |
| Storage conditions | RT (15-25°C) | |
| Shelf Life of Different Surfaces | | |
| ibiTreat, Glass Bottom, ESS | 36 months | |
| Collagen, Poly-L-Lysine | 18 months | |

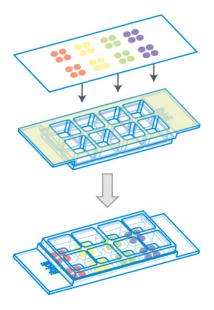
Handling and Assembling

Assemble the sticky–Slides with a convenient bottom material, matching your experimental needs. Use our Clamp for sticky-Slides for a comfortable assembling (ibidi, 80040).

- Prepare your sample and/or bottom material.
- Remove the protection film by using sterile tweezers.
- Optionally for channel sticky–Slides, place your sample into the channel.
- Mount bottom and sticky–Slide with some pressure. Press well until the bottom is sealed. For best results use our Clamp for sticky–Slides (ibidi, 80040).
- Incubate at 20-40°C for best results.
- Conduct your experiment.







The adhesive strength strongly depends on temperature and time. Best results are achieved by storing the assembled Slides over night at 20-40°C. Anyhow, sticky–Slides are not leaky immediately after assembling.

sticky–Slides can be removed from the substrate by dipping them into Acetone over night in an appropriate glass container (e.g. a beaker). Please keep in mind that Acetone might be harmful to your used substrate. Once removed sticky–Slides cannot be reused.

Surface Compatibility

sticky–Slides are compatible with all flat, clean, dust–free, fat–free surfaces like glass, plastic, metal, silicium or electrode structures. sticky–Slides can be assembled with wet surfaces (protein–free, aqueous solutions like water or PBS buffer). Dusty or fatty surfaces like wax foils or similar surfaces are not compatible. Please test your specific surface in your lab with free samples from www.ibidi.com.

Best results are achieved when flexible substrates like plastic sheets or coverslips are used. Rigid glass slides or metal surfaces are also possible to use but need more pressure to seal.

Seeding Cells

 Trypsinize and count cells as usual. Dilute the cell suspension to the desired concentration. Depending on your cell type, application of a 4–9 × 10⁴ cells/ml suspension should result in a confluent layer within 2–3 days.

- Apply 300 µl cell suspension into each well of the Slide. Avoid shaking as this will result in inhomogeneous distribution of the cells.
- Cover reservoirs with the supplied lid. Incubate at 37°C and 5 % CO₂ as usual.

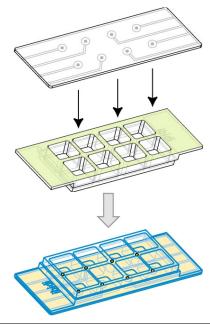
Undemanding cells can be left in their seeding medium for up to three days and grow to confluence there. However, best results might be achieved when the medium is changed every 1–2 days. Carefully aspirate the old medium and replace it by $300~\mu$ l/well fresh medium.

Immersion Oil

Immersion oil compatibility depends on the used substrate.

Applications

The sticky–Slide 8 Well provides a common open well format which is best suited for maximum sample access, e.g. when cells have to be seeded onto a titanium implant material.



Solvents for Fixation, Staining and Other Purposes

The sticky bottom material and the slide material are compatible to Methanol, acids, alkalis, PFA, DMSO, and silicone oil. Please keep in mind that these substances may be harmful to the used substrate. Acetone is not compatible with the sticky material so it can be used to detach slide and substrate after use.



sticky-Slide 8 Well





sticky-Slide 8 Well

Instructions

Ordering Information

The sticky–Slide technology is available with different slide formats. Please see the table below for choosing your sticky–Slide.

sticky-Slides

| Cat. No. | Description |
|----------|--|
| 80828 | sticky–Slide 8 Well: sterilized |
| 80328 | sticky–Slide VI ^{0.4} : sterilized |
| 80608 | sticky-Slide Chemotaxis: sterilized |
| 81128 | sticky–Slide I ^{0.1} Luer: sterilized |
| 80168 | sticky–Slide I ^{0.2} Luer: sterilized |
| 80178 | sticky–Slide I ^{0.4} Luer: sterilized |
| 80188 | sticky–Slide I ^{0.6} Luer: sterilized |
| 80198 | sticky–Slide I ^{0.8} Luer: sterilized |
| 10812 | Coverslips for sticky–Slides: #1.5H (170 μ m \pm 5 μ m) D 263 M, Schott glass, 25 mm \times 75 mm, unsterile |
| 10813 | Coverslips for sticky–Slides Uncoated: #1.5 polymer coverslip, 25 mm × 75 mm, unsterile |
| 10814 | $\textbf{Coverslips for sticky-Slides ibiTreat: } \$1.5 \text{ polymer coverslip, tissue culture treated } 25 \text{ mm} \times 75 \text{ mm, unsterile } \$1.5 \text{ polymer coverslip}, \texttt{tissue culture treated } 25 \text{ mm} \times 75 \text{ mm, unsterile } \$1.5 \text{ polymer coverslip}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm}, \texttt{tissue culture treated } \$1.5 \text{ mm} \times 75 \text{ mm} \times 75 \text{ mm}, tissue culture t$ |

Clamp for sticky-Slides

| Cat. No. | Description |
|----------|--|
| 80040 | Clamp for sticky-Slides |
| 80041 | Adapter for sticky–Slide 8 Well: |
| 80042 | Adapter for sticky–Slide I Luer: |
| 80043 | Adapter for sticky–Slide VI ^{0.4} : |
| 80044 | Adapter for sticky-Slide Chemotaxis: |

For research use only!

Further technical specifications can be found at www.ibidi.com. For questions and suggestions please contact us by e-mail info@ibidi.de or by telephone +49 (0)89/520 4617 0. All products are developed and produced in Germany.

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